

Prospects, promise and problems on the road to effective vaccines and related therapies for substance abuse

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This review addresses potential new treatments for stimulant drugs of abuse, especially cocaine. Clinical trials of vaccines against cocaine and nicotine have been completed with the generally encouraging result that subjects showing high titers of antidrug antibody experience a reduction in drug reward, which may aid in cessation. New vaccine technologies, including gene transfer of highly optimized monoclonal antibodies, are likely to improve such outcomes further. In the special case of cocaine abuse, a metabolic enzyme is emerging as an alternative or added therapeutic intervention, which would also involve gene transfer. Such approaches still require extensive studies of safety and efficacy, but they may eventually contribute to a robust form of *in vivo* drug interception that greatly reduces the risks of addiction relapse.

KEYWORDS: adenovirus gene transfer vector • antidrug immunoglobulin • butyrylcholinesterase • clinical trial • cocaine abuse • cocaine hydrolase • metabolism-based therapies • monoclonal antibody • vaccine

According to the 2010 National Survey of Drug Use and Health, over 22 million Americans have substance dependence or substance abuse problems [10]. Many stimulants do not directly target a clearly defined chemoreceptor, but act on transporters to increase synaptic levels of dopamine, norepinephrine and serotonin in brain reward centers. For this reason, successful treatment options with receptor antagonists have been difficult to identify. One strategy for overcoming these obstacles and reducing risks of relapse after initial rehabilitation is based on intercepting drugs of abuse before they reach brain reward centers. The major therapeutic candidates to implement such an approach are antibodies and catalytically efficient metabolic enzymes, both of which may be considered pharmacokinetic antagonists of their target drug. A number of vaccines have been developed to elicit high-affinity antibodies against different stimulants with high liability for abuse. These vaccines and their clinical efficacy have been discussed in several recent reviews [1–7], but the pace of research remains rapid and the subject merits continued attention. A different but related means of drug interception involves accelerating metabolic clearance to the

point where it can also affect the levels of drug reaching the brain. This idea arose not long ago, and, for reasons the authors will discuss, appears applicable only to a single agent of abuse, namely cocaine. This review will consider progress and obstacles on the path to developing and implementing vaccine and metabolism-based therapies optimal for the treatment of cocaine abuse. The authors will also explore the potential for the two interception approaches to reinforce each other in an additive or even synergistic manner.

Antibodies & vaccines against drugs of abuse

In recent years, the therapeutic scope of vaccines has been broadened by the realization that toxic drugs of various kinds can be prevented from reaching their sites of action if the recipient has been vaccinated against a closely related chemical structure. When it comes to addictive substances, it has been proven feasible to generate substantial titers of high-affinity antibodies against psychoactive small-molecule haptens conjugated to appropriate carrier proteins such as keyhole limpet hemocyanin, cholera toxin [8] and tetanus toxoid [4].

It is recognized that any drug–haptent conjugate will elicit a strong antibody response to the carrier protein as well as the intended target, but these responses do not interfere with the production of antidrug immunoglobulins and anticarrier antibodies are regarded as innocuous, although they can sometimes affect production of the desired antihaptent antibodies [9]. The nature of the carrier protein is particularly important in determining the ultimate titer of antidrug antibodies. One new conjugating agent showing great promise in this regard involves capsid proteins from killed adenovirus [10].

Drug–haptent conjugated vaccines have proven able to reduce the physiological and behavioral responses to their target molecules when tested in experimental animals, typically rodents, but also primates. Encouraging results have been obtained with vaccines to cocaine, heroin, nicotine and methamphetamine, among other widely abused stimulants [11–14]. Thus, several studies have demonstrated reduced total drug levels in plasma or brains of rodents after passive immunization or after a good vaccine response with high titers of specific drug antibody [15–18].

Such vaccines are capable of eliciting antibodies in molar quantities approximately commensurate with the plasma drug levels attained after typical ‘rewarding doses’. This point raises both a technical and substantive issue. First, although the classic immunology literature typically deals with antibody titers in terms of thresholds for detection, molar values are more relevant to questions of balance between drug molecules and ligand binding sites on an immunoglobulin. Second, there is the practical question of how many drug binding sites are needed when the objective is to weaken or eliminate drug actions in the CNS, recognizing that each antibody has two binding sites and, in theory, can bind two drug molecules. In the absence of experimental data, a one-to-one ratio between binding sites and drug molecules would seem a reasonable goal. Published studies, however, indicate that drug-binding immunoglobulins with high affinity affect reward-driven behavior noticeably even when, in terms of local concentration, they are somewhat less abundant than the drug itself [19,20].

Since a sudden rise in brain drug concentration is regarded as key to drug reward [21,22], antibody-induced blunting of reward effects is often ascribed to a broadening of the ‘drug pulse’ created by rapid intake (i.e., reduced rate of rise in drug concentration). Indeed, turbulent flow allows the drug to interact with antibodies while passing through the vasculature. IgG distributes throughout the extracellular water compartment, at levels somewhat lower than in blood. Since this compartment is several times larger than plasma volume, the total pool of antibody available to adsorb the drug can substantially exceed the total drug dose. Under this condition, specific antidrug IgG can reduce the concentration of free drug and the rate at which the drug concentration rises in the brain. Without a sophisticated dynamic model, the exact consequences are unpredictable. However, this line of reasoning suggests that such effects can be physiologically important.

Advantages & problems with antidrug vaccines

The vaccine approach to treating drug abuse has two great advantages. First, the basic technology is well understood, robust,

cost-effective and easy to accomplish; and second, the risks of treatment are relatively minor, both in reality and in the public perception. In the authors’ view, the chief problems are these: individual responses can be highly variable (for genetic and other reasons); responses in many human subjects do not eliminate drug reward entirely and may not be sufficient therapy in themselves; there are no ready means of insuring that a large proportion of the antibodies generated are optimal for drug binding; sustained therapy requires booster immunizations and, therefore, continued commitment to remaining drug free; any given level of antidrug antibody can, in principle, be saturated by a large or repeated drug dose; a subject taking increased doses, especially of cocaine, may accumulate toxic intermediates that damage vulnerable tissues such as the liver or heart (recent work in the authors’ laboratories to model such effects will be discussed later). Most of these problems are amenable to further advances in vaccine technology, including better adjuvants and haptent conjugates, and perhaps novel ways of generating and delivering immunoglobulins with ideal properties toward the target agent(s). Improvements can be expected on a number of fronts as the basic science develops. Meanwhile, progress is occurring at the clinical level. It can be argued that existing vaccines already tip the balance between cost (in risk and effort) and drug reward. Further improvements could well cause a noticeable reduction of drug seeking and drug intake.

Clinical trials

Clinical trials have been successfully completed in vaccines for cocaine [23] and nicotine [24,25]. The results indicate partial therapeutic effects with both target drugs. Such effects are, of course, related to the antibody titers achieved. Because these tend to vary substantially among individuals, the overall compiled results are modest and, in the case of the cocaine vaccine, not statistically significant. A major encouragement from the cocaine vaccine study by Kosten and colleagues was a statistically significantly increased frequency of drug-free urine [8]. This outcome was impressive given that the participants were regular users with no declared motivation to quit. Another striking feature of this study was the apparent failure of participants to compensate for a hypothesized reduction in perceived reward value of cocaine. Thus, close monitoring of the urine samples revealed no pattern of increase in cocaine metabolites, which would have accompanied a rise in drug intake. Studies at Baylor College of Medicine (TX, USA) showed that some subjects developed IgM anticocaine antibodies in response to prior recreational exposure and they responded significantly less well to the cholera toxin–conjugate cocaine vaccine [26]. The reason for antibody development is not known, but it is possible that reactive cocaine metabolites combined *in vivo* with host proteins to form haptent–carrier complexes. One lesson is that future trials must be designed explicitly to stratify participants in terms of anticipated levels of antidrug–antibody response. This would allow data forwarded to the US FDA or international regulatory bodies to meet expectations of success. It also seems appropriate in future trials to exclude subjects with high titers of pre-existing anticocaine IgM, which may arise by mechanisms similar to those operating in the Baylor vaccine trial.

Monoclonal antibodies

New ideas and technologies are emerging to enhance the power of antidrug vaccines or complement them in synergistic ways. In particular, recent work has yielded notable successes in the form of monoclonal IgG antibodies (mAbs) with very high affinities for target agents such as amphetamine, cocaine and nicotine [14,17,27,28]. Such antibodies are clearly able to affect reward-driven responding in rodents, although not entirely in the fashion one might expect. For example, with a humanized, mouse-derived mAb to cocaine, Norman and colleagues found that a large dose delivered by intravenous injection (120 mg/kg) led to a threefold increase in the 'priming threshold' in rats trained for cocaine self administration [27]. This effect can be viewed as a sign of reduced reward value for the amount of cocaine delivered. In addition, there was a modest dose-dependent increase in cocaine consumption rates, which was interpreted as an attempt to attain the same final reward level that was being attained before antibody administration. These findings imply that the mAb was affecting cocaine delivery to brain reward centers, but to a modest and surmountable extent. One might hope for a complete cessation of responding if interception were maximally effective. This goal may actually be achievable with higher doses of the mAb, or one with greater affinity for the target. At present, however, even a partial effect of this nature could help individuals and benefit communities.

A chief obstacle in implementing mAbs for drug abuse (passive immunotherapy) is the need for large amounts of purified IgG, in addition to the cost and inconvenience of repeated injections, or the vagaries of sustained release preparations, mini-pumps and other means of sustained delivery. Drug-specific mAbs might well serve to reverse drug overdose rapidly in emergency situations. However, passively administered antibodies can only remain in the system for a short period of time, assuming normal metabolic clearance. Although considerable therapeutic impact could be gained by selection and protein engineering for ideal drug-binding properties and *in vivo* behavior, these advantages are compromised when long-term therapy requires the patient to make a series of affirmative decisions to continue with a treatment plan that offers no direct relief or reward. One could argue that the basis for successful treatment should be an approach that is truly 'vaccine-like' in the sense that an initial decision is followed by steps that are inherently long-lasting, with no need for frequent interventions to maintain them. In this respect, conventional vaccination with the best available carriers and adjuvants lies in the border zone. That is to say, such treatments can provide high titers of antidrug antibodies over periods much longer than the half-life of circulating IgG molecules, but several revaccination or booster injections per year may be required to sustain the immune response. This state of affairs can work well for individuals with an appropriate level of motivation. Nonetheless, it is a reason to explore other means for long-term delivery of drug-intercepting proteins.

Gene transfer of antidrug proteins

Gene transfer can deliver a refined and reproducible therapeutic protein with the durability and compliance-friendly aspects of

vaccination. After a hiatus of more than 10 years following the unfortunate outcome of an early clinical trial of protein therapy based on gene transfer [29], this approach is re-emerging as an option for a number of chronic conditions, including clotting disorders [30], immunodeficiency [31], inherited blindness [32] and cancer [33]. Substance abuse and addiction may also prove amenable to gene transfer, in this case by delivering 'interceptor molecules' that prevent drug access to brain targets.

The characteristics of ideal gene therapy for any nonfatal illness should be, in approximate order of importance: acute safety (no serious immediate adverse effects); chronic safety (no immunological disturbance or disruption of oncogenes in cells harboring delivered DNA and producing transgene product) and high therapeutic impact. To aid in recovery from addiction, one also needs to sustain adequate levels of therapeutic protein for the period with maximal risk of relapse (which can last 2 years or more). Since addiction is not immediately life-threatening, these requirements raise the bar for treatment with viral vectors. However, to argue the positive case, recognizing the huge burden that drug addiction imposes on individuals and families, an effective therapy with some degree of risk does seem appropriate.

Despite lingering concerns regarding viral gene transfer, its potential for delivery of 'ideal' monoclonal antibodies is well recognized [34,35] and has recently been tested in animal studies on viral transduction of an antinicotine IgG. Using an AAVrh.10 vector that expressed full-length antibody derived from the Fab fragment of antinicotine mAb NIC9D9, Hicks and colleagues were able to shield mice brains from systemically administered nicotine, reducing tissue drug concentration approximately sevenfold as compared with unprotected mice [36]. This effect was adequate to block nicotine-stimulated increases in blood pressure, heart rate and locomotor activity. Moreover, the antibody persisted undiminished at high titers for 18 weeks. In fact, the levels of antinicotine IgG achieved after injections of 10^{11} viral vector gene copies (moderate viral load) rose to 1000 $\mu\text{g/ml}$, equivalent to approximately 150 μM concentration of drug-binding sites. This was far above what could be expected after classical vaccination, and approximately tenfold higher than the nicotine plasma concentration observed in human volunteers immediately after smoking a high-nicotine brand of cigarettes [37]. Therefore, the highly effective action in mice could have been expected and it may well translate to human application. The same research group achieved similar success with viral delivery of an anticocaine antibody, which suppressed cocaine-induced locomotion for months [38].

Immunological approaches through gene transfer along these lines offer several advantages. First, the molecular properties of the therapeutic can be defined and refined ahead of time. Second, long-term cost should be far lower, as there is no need for repeated large-scale culture fermentations and purification. Third, a single treatment should suffice to generate therapeutic levels of antibody long enough to stabilize abstinence – that is, more than a year and perhaps indefinitely. The duration of viral transduction depends almost entirely on the nature of the vector. Truly permanent transduction requires insertion of vector DNA into host chromosomes, which can be accomplished with

retroviral vectors based on lentivirus or similar agents [39]. As is well known, such vectors pose a small but nonzero risk of disrupting oncogenes and setting the stage for tumor formation [40]. Safer vectors include those based on adenovirus, including the classic E-1-deleted vectors [41] along with the more refined helper-dependent adenovirus (HDAd) [42] and adeno-associated virus (AAV) [43] constructs. These noninserting vectors sustain transduction by persisting as nonreplicating episomal elements in the host cell cytoplasm. When the cell divides, however, vector DNA is lost or diluted. Therefore, transgene transduction depends on the lifespan of the target cell, which is unlimited in neurons, cardiac and skeletal muscle cells, quite brief in most epithelia and smooth muscle cells, but moderately long in liver cells (1–3 years depending on species) [44,45].

Drawbacks & obstacles to gene transfer

One major present drawback is a persistent concern for safety, which is well-founded and deserves attention. A key risk factor appears to be the brief but intense innate immune response to viral capsid proteins [46]. Much research has concentrated on this issue and how to minimize or avoid it [47,48]. Deletion of multiple viral genes while preserving transduction capability is one important step, as in the engineered helper-dependent adenoviral vectors [49] or the vectors based on naturally helper-dependent adeno-associated viruses [50,51]. These modified agents typically have no ability to express viral proteins *in vivo* and thus, in the longer term, may remain hidden from immune surveillance [52]. On the other hand, to deliver an engineered DNA payload, viral particles must first attach to the plasma membranes of host cells that can translate the genetic information into a protein product. This function requires packaging of the functional DNA within a coat that protects it from dispersal and metabolic attack, selectively binding surface targets that are characteristic of the selected host cells and stimulating endocytosis of the delivered DNA. At the present state of vector technology, the most efficient means of accomplishing this objective uses artificial protein coats derived from the shell of a relatively benign ‘helper virus’. The immune system still recognizes these coats, which remain able to trigger a seriously adverse response. However, such responses are typically short-lived. They are also dose related and can be minimized by delivering a smaller load of vector, albeit at the price of obtaining less transgene product.

Fortunately, transient immunosuppression with anti-inflammatory steroids, such as dexamethasone, will reduce host immune responses to viral vectors and lessen subsequent toxicity [53] while also prolonging transgene expression [54]. Often, immunosuppression is also required for adequate levels of therapeutic protein. This is especially true in animals subjected to procedures creating inflammatory stress (e.g., implantation of multiple cannulae), which can impair subsequent virally mediated transgene expression [GAO Y, BRIMIJOIN S, UNPUBLISHED DATA]. In higher organisms, especially nonhuman primates and humans, the acute immune response to vectors is very strong, associated with a sharp rise in cytokines such as IL-6, while immunosuppression is more difficult and less effective [55]. Progress in this area is critical to future success.

Accelerated metabolism

As noted initially, accelerated drug metabolism is developing as an alternative way of reducing cocaine access to the brain, a key step in the reward process that generates and maintains addictive behaviors. This approach is tenable only because of butyrylcholinesterase (BChE), a single plasma enzyme that converts cocaine into virtually inactive metabolites, benzoic acid and ecgonine methyl-ester [56]. By contrast, the metabolism of amphetamines, nicotine and many other stimulant drugs depends on cytochrome-p450-dependent pathways in the liver [57,58]. These pathways require intact electron transport chains that only operate in an intracellular environment and cannot readily be enhanced by an exogenous enzyme. As a potential therapeutic agent, BChE appears very safe. Like acetylcholinesterase (AChE), it hydrolyzes acetylcholine; however, it plays little, if any, role in cholinergic transmission or any other known physiological process. Also, as a natural constituent, BChE accompanies every plasma transfusion without discernible effects. In view of these facts, in 1997 Gorelick proposed using human BChE to treat cocaine overdose [59]. Because natural BChE is not highly efficient with cocaine, Lockridge and collaborators tried site-directed mutagenesis and obtained a BChE with a fourfold improved catalytic activity [60]. This process, aided by computer-based modeling of cocaine docking and catalysis continued in other laboratories, including the authors and that of Zhan, led to a series of ‘rationally designed’ BChE mutants with increasing catalytic activity against cocaine [61–65]. The key enzyme yardstick is ‘catalytic efficiency’, defined as the ratio between k_{cat} (molecules of substrate hydrolyzed per min, per active site) and K_m (substrate concentration at half-maximal velocity). Unless a drug saturates the binding site (an improbable condition), an efficient enzyme will outperform others with higher catalytic power (larger k_{cat}) but disproportionately higher K_m . Compared with natural BChE, the optimal mutant cocaine hydrolase (CocH) is less efficient with acetylcholine but 1300-fold more efficient with cocaine [65].

The advent of catalytically efficient CocH led to the finding that an enzyme can not only prevent cocaine toxicity in animal models, but also reverse it. In fact, CocH is able to abort lethal cocaine-induced seizures in rats and reliably rescue them even when delivered after major convulsions have already begun [66]. Even more interesting are findings indicating that pretreatment with CocH prevents cocaine-primed reinstatement of cocaine-seeking behavior in rats trained on cocaine self-administration [67]. This outcome pointed toward a possibility of using enzymatic therapy to aid recovering addicts avoid relapse into drug taking. Encouragingly, unmodified BChE has proven remarkably benign in studies undertaken by the US Department of Defense for prophylaxis against chemical warfare agents. In fact, rodents showed no ill effect after treatment with BChE in doses that raise plasma levels hundreds of fold [68–70]. Human subjects given gram quantities of the enzyme also experienced no perceptible changes, subjective or objective. In this sense, BChE seems physiologically inert. One can speculate that it evolved for metabolic disposition of bioactive esters in the diet. In the authors opinion, however, direct administration, even of a safe and effective esterase, is still

problematic for the chronic treatment needed with addiction. Expensive and highly purified protein would have to be given frequently and in large amounts. Even slow-release preparations would often need renewal. Costs could be exorbitant and patient compliance may be problematic. These facts led us to consider delivering BChE-based CocH by therapeutic *in vivo* gene transfer, either as a stand-alone treatment for relapse prevention, or as part of a combination therapy with an anticocaine vaccine.

CocH gene transfer with helper-dependent adenoviral vector

To date, several animal studies have been completed with a view towards assessing the safety and efficacy of CocH gene transfer. Early studies were based on a first-generation E-1 deleted adenoviral vector. The results in rats showed very high levels of transgene expression, elevating CocH activity in plasma by an average multiple of 50,000 [71]. However, such levels were only sustained for a few days, being limited by a strong host response to transgene and probably to transduced hepatocytes as well. To achieve more stable transduction, the authors have utilized two newer-generation vectors: first, a HDAd based on the constructs developed by Parks, Mitani and Kochanek [42,72,73]; and more recently, an adeno-associated viral vector [74,75].

The viral coat of the HDAd vector is similar to the older vector, as is the initial immune response. However, lacking DNA for all viral proteins, HDAd does not provoke sustained immunological reactions. This vector is a clear advance and has an impressive ability to sustain long-term transduction in the liver. Thus, Ng and collaborators obtained high circulating levels of human α -antitrypsin for over 1 year in baboons [49] and lifetime expression (~2.5 years) of the *ApoE* gene in genetically deficient mice [76]. The authors strategy also targets the liver as the locus for gene transduction, given that this organ is the natural site of BChE synthesis and release into plasma, and that its high blood flow ideally suits it for metabolic surveillance. In addition, hepatocyte lifespan, both in rodents and humans, should permit effective transduction of the enzyme for a considerable period of time.

Current work has demonstrated that gene transfer of CocH with a liver-directed HDAd vector can sustain very high levels of circulating enzyme in rodents. Specifically, the authors obtained 1000-fold increases of cocaine hydrolyzing activity for at least 6 months in rats [67]. In separate and ongoing studies, mice are showing substantial hydrolase transduction 18 months after initial treatment [GAO Y, GENGL, BRIMIJOIN S, UNPUBLISHED DATA]. Therefore, continued testing is focused on hepatotropic viral vectors incorporating liver-specific promoters. Signs of toxicity have been absent at the doses used (up to 10^{13} particles per mouse, delivered through the tail vein), which elevated circulating CocH activity by a factor of approximately 1,000,000 [77]. In particular, blood monitoring for liver enzymes has detected no elevation (implying continued integrity of hepatocytes), and liver tissue has appeared entirely normal in postmortem histological sections. Treated mice show normal behavior patterns, normal levels of spontaneous activity and weight gain, and when challenged with doses of cocaine that are hepatotoxic or lethal in unprotected animals, they

show dramatically reduced toxicity or none at all [77]. Rats given a single vector injection also showed no sign of adverse effects. However, during the entire 6-month period of observation, the treatment blocked the drug-seeking 'reinstatement behavior' usually triggered by cocaine exposures [67]. Somewhat surprisingly, a retrospective analysis of the data revealed that this failure of cocaine-primed reinstatement persisted even in the few rats that lost CocH expression after 3–4 months. This effect was specific to cocaine. Thus, at 6 months, the same rats showed robust drug-seeking after a priming injection of amphetamine, a stimulant not metabolized by CocH. A possible explanation for persistent loss of cocaine-primed reinstatement behavior is that vector-treated rats unlearned the association between cocaine intake and a drug-related reward. Wise and colleagues have provided convincing evidence that organisms associate drug rewards with 'interoceptive cues' from peripheral chemoreceptors [78]. These cues are relayed to the brain by fast neural pathways and initiate a release of glutamine in the ventral tegmental area. The glutamate 'spike' is followed by a wave of dopamine release caused by the drug acting directly on central reward nuclei. In subjects with abundant CocH, the interoceptive cues may persist but unaccompanied by a central reward. Multiple repeats of this experience might eventually extinguish drug-seeking behavior triggered by drug exposure. If the metabolic products of cocaine, namely benzoic acid and ecgonine methyl-ester, had mildly aversive properties, this effect could be even stronger. The authors have no answer to this question as yet but consider it worth exploring.

Combining gene transfer & vaccination

Anticocaine vaccine and CocH gene therapy deserve to be considered together because they are different means to the same end: preventing an addictive drug from reaching its principal site of action. In addition, these two different modes of drug interception are compatible with one another and should be mutually reinforcing. Several factors contribute to this potential. Vaccination can easily generate specific antibody titers in the low micromolar range, implying a capacity to bind a major fraction of a typical reward-level drug dose. Since antibody binding occurs almost immediately [79], an initial pulse of drug is blunted. However, a quickly repeated or larger dose can still exert central effects because antidrug antibodies are limited in abundance and therefore saturable. In comparison with vaccination, gene transfer of an enzyme is likely to generate fewer drug binding sites, with lower affinity. However, catalytic action can eliminate virtually any amount of drug and restore a normal state within minutes. In an *in vitro* model, when CocH was added to a 1 μ M solution of cocaine >90% bound with nanomolar affinity to anticocaine IgG, the enzyme destroyed 98% of all the drug within 90 s [80]. This reaction is easily fast enough for physiological relevance.

More stringent tests of CocH have been carried out in animal models. In mice challenged with large doses of cocaine, at or near the LD_{50} (i.e., 100–120 mg/kg, intraperitoneally), single treatments with anticocaine antibody (8 mg/kg) or CocH (1 mg/kg) did not reduce motor dysfunction and provided only modest protection of liver structure. Given together, however,

the same treatments effectively prevented all these toxic actions of the drug [77]. Similar cooperativity was noted with regard to cocaine-induced locomotor activity in rats [81]. The subjects were given keyhole limpet hemocyanin–norcocaine vaccine approximately 6 weeks before testing and a booster injection at 3 weeks, or they received HDAd vector with CocH cDNA approximately 2 weeks before testing. Under the experimental conditions, individual treatments reduced cocaine stimulation modestly, while combined treatments provided much greater reduction. To prove that such results reflect truly synergistic actions will require rigorous investigation of a broader range of treatment levels, along with isobolographic analysis. Meanwhile, the benefits are at least additive. Therefore, anticipating future therapy of human drug users, the authors envisage dual treatments by vaccination and CocH gene transfer. The primary goal would be to provide more robust protection from addiction relapse than could be obtained from either treatment alone. In addition, combined treatments might permit reduced vector dosages and less frequent booster immunizations, resulting in increased safety, lower costs and reduced inconvenience.

Potential problems & pitfalls with CocH gene therapy

A general limitation of gene therapy, apart from the risks of toxicity already discussed, is related to acquired immunity directed against the specific viral antigens presented by the particular gene-transfer vector to be used. This problem arises in two forms. First, if a subject has previously been exposed in daily life to a virus of the same serotype as the vector, active immunity will already be in place and the initial transduction will fail. This event may pose some risk to the recipient and it will block future attempts at gene transfer with that vector. For the same reason, when a subject experiences successful transduction but needs to prolong it, readministering the same vector will be ineffective. Therefore, pretreatment screening for viral protein antibodies will be essential. Fortunately, however, work in animal models indicates that ‘serotype switching’ or the use of a vector based on a different virus is an effective solution [82].

Another question is whether CocH transduction might arouse immune responses that curtail the therapeutic effect. The sustained, high levels of cocaine hydrolyzing activity in the liver and plasma after viral gene transfer in rats implies that the mutated human BChE was tolerated by their immune system, at least to a degree. Nonetheless, looking further into the issue, the authors tested directly for anti-BChE IgM or IgG in rat blood samples at multiple intervals after transduction. None were detected at 3, 20 and 30 weeks. A few animals tested weakly positive at 50 weeks, but CocH serum half-life (measured by recovery from irreversible inhibition) was not shortened [83]. The authors conclude that CocH is only weakly antigenic in this species.

The long duration of HDAd-mediated CocH transduction is partly due to the lack of encoded DNA for viral proteins. This advantage is shared by AAV-based vectors, the most commonly used gene-transfer agents in recent clinical trials. Ongoing studies in the authors laboratories are investigating AAV for CocH transduction in mice. Mice appear somewhat less tolerant to

CocH than rats. When transduced by low-dose AAV, type 2/8, mice develop antibodies to human BChE and lose most of their transgene expression within 2 weeks [GAO Y, BRIMIJOIN S, UNPUBLISHED DATA]. Immune reactions to native BChE should not occur in human subjects other than those rare individuals with a null gene mutation [84,85]. However, the occurrence of anti-BChE antibodies in rats and mice raises the question of whether CocH, as a BChE mutant, could be weakly antigenic in humans. The authors hypothesis is ‘no’, because all mutations are confined to the catalytic gorge and at least partly hidden from immune surveillance. This hypothesis deserves to be tested before working toward an eventual human trial. For that purpose, comparable mutations were made in mouse BChE, and immune responses were examined after transduction with HDAd or AAV. Unpublished results in both cases indicate enzyme persistence for months with complete absence of antibodies to the transgene product. Because mutating the key sites in a conspecific enzyme does not confer immunogenicity, it seems likely that the equivalent mutations in human BChE will also be immunologically silent.

One theoretical concern regarding greatly elevated plasma CocH is that this BChE-derived enzyme retains substantial ability to hydrolyze the cholinergic neurotransmitter acetylcholine [86]. In actuality, the likelihood that CocH will cause problems with cholinergic neurotransmission is remote. Extensive prior research has shown little physiological impact of exogenous BChE. Thus, studies to evaluate human BChE as a prophylactic treatment for exposure to chemical warfare agents uniformly failed to detect physiological disturbances in rats, guinea pigs and primates, even after administering near-gram quantities that raised plasma enzyme levels 50–100-fold [68–70,87–89]. This impressive evidence for safety and the absence of cholinergic dysfunction is counterintuitive but not illogical. There are at least two reasons. First, mammalian blood contains approximately equivalent molar concentrations of AChE (in plasma and red cells) and BChE (plasma only), but the former is over twice as catalytically efficient with acetylcholine [90,91]. Hence, a tenfold increase in circulating BChE only triples hydrolysis capability. More important, motor synapses are all packed with cholinesterase at levels thousands of times above those in general circulation or extracellular water. Elegant morphometric studies by Anglister and colleagues provided a reliable basis for quantifying actual AChE concentrations within the synaptic cleft of the neuromuscular junction [92]. With a snake neurotoxin serving as a highly selective and high-affinity probe for this enzyme (¹²⁵I-labelled fasciculin) these investigators established that the density on the synaptic membrane was 5×10^{19} catalytic subunits per ml, or roughly 10^{-4} M. By contrast, published values indicate the concentration of BChE subunits in normal mouse serum to be approximately 2×10^{-9} M. Therefore, considering relative catalytic efficiencies, it would require at least a 100,000-fold increase in plasma BChE levels to match the ACh-hydrolyzing capacity of motor synapses. Thus, even the upper limits of plasma BChE expression are highly unlikely to affect transmitter levels or duration at motor synapses. Cholinergic synapses in the brain are also unlikely to be disturbed as they are well insulated from

circulating hydrolase by virtue of the blood–brain barrier, which excludes BChE [93]. Among the few remaining areas for concern are the nicotinic synapses in the peripheral autonomic ganglia and, perhaps, muscarinic synapses at end organs innervated by parasympathetic nerves. Investigations of cardiovascular function in vector-treated mice are presently underway.

Conclusion

There is exciting potential for broad application of antidrug vaccines as one component of a comprehensive approach to managing drug addiction, aiding in recovery and preventing relapse. Vaccines that generate effective drug-binding IgG antibodies at high titers are well along the road to clinical application for treating both nicotine and cocaine abuse. The feasibility of long-term gene transfer delivery of even more effective monoclonal antibodies for the same drugs has recently been established. Finally, in the case of cocaine, metabolic elimination of the drug by highly efficient engineered enzymes holds promise as an alternative or complementary strategy for intercepting the stimulant before it reaches brain reward centers.

Expert commentary

Continuously improving technology should be applied to increase the effectiveness of classic vaccines against drugs of abuse. The key to success will be increased titers of high-affinity antidrug antibodies. Promising avenues to explore include conjugation of drug haptens with more highly immunogenic carrier proteins, such as those derived from adenovirus capsids. Delivering optimized monoclonal antidrug antibodies by gene transfer over extended periods without the need for repeated interventions is another idea well worth exploring. Issues of safety and efficacy with that approach remain significant but are probably surmountable. As the demonstrated and perceived safety of gene transfer improves, this technology can also be applied to deliver drug-metabolizing enzymes. At present, the obvious choice for a therapeutic antidrug enzyme is a CoCH based on natural human plasma cholinesterase. Besides having therapeutic potential on its own, such an enzyme

is likely to complement and magnify the effects of antcocaine vaccines.

Five-year view

One can safely predict incremental advances in antidrug vaccine technology as the choice and design of haptens and carrier proteins are refined. The authors speculate that in 5 years' time, there will have been strongly positive outcomes from clinical trials of several treatment categories touched upon in this review with regards to cocaine and nicotine abuse. In particular, classic vaccines with more effective carrier proteins and adjuvants will have proved able to elicit higher levels of antidrug IgG than previously seen, with more than a third of the subjects attaining levels associated with a marked reduction of abuse liability. Second, clinical gene transfer of a 'humanized' high-affinity monoclonal antibody to nicotine or cocaine will have succeeded in generating still higher levels of antidrug antibodies and greater *in vivo* drug antagonism, accompanied by limited and tolerable side effects during the initial stages of transduction. Third and finally, a clinical trial of viral gene transfer of CoCH will be in the process of demonstrating safety and a degree of efficacy that, if not adequate in itself, could greatly amplify the therapeutic results of an antcocaine vaccine.

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Key issues

- Vaccines have been developed against multiple addictive substances including cocaine, nicotine, amphetamine and heroin, with the aim of reducing abuse liability and aiding abstinence.
- Such vaccines are based on the conjugation of drug haptens to immunogenic carrier proteins.
- Existing vaccines, in clinical trial or current use, are moderately effective at best.
- Drug vaccines under development utilize more intensely immunogenic carrier proteins, such as coat proteins from killed adenovirus, to generate higher titers of antidrug antibodies.
- Under investigation as a possible alternative to conventional vaccines is gene transfer of monoclonal antidrug antibodies engineered for optimal affinity.
- Gene transfer of drug-metabolizing enzymes is an alternative 'vaccine-like' approach to treatment of substance abuse.
- Recent animal studies involving therapeutic gene transfer of cocaine hydrolase validate the concept of accelerated drug metabolism as an alternate means of reducing cocaine's abuse liability.
- Current research suggests that a combination of the immunologic approach (vaccines and antibodies) and the metabolic approach (gene transfer of enzymes) is likely to be particularly effective.

References

Papers of special note have been highlighted as:

• of interest

•• of considerable interest

- 1 Shen XY, Orson FM, Kosten TR. Vaccines against drug abuse. *Clin. Pharmacol. Ther.* 91(1), 60–70 (2012).
- **Up-to-date survey of the current status of antidrug vaccines.**
- 2 Shen X, Kosten TR. Immunotherapy for drug abuse. *CNS Neurol. Disord. Drug Targets* 10(8), 876–879 (2011).
- 3 Shen X, Orson FM, Kosten TR. Anti-addiction vaccines. *F1000 Med. Rep.* 3, 20 (2011).
- 4 Orson FM, Kinsey BM, Singh RA, Wu Y, Gardner T, Kosten TR. Substance abuse vaccines. *Ann. N. Y. Acad. Sci.* 1141, 257–269 (2008).
- 5 Hatsukami DK, Stead LF, Gupta PC. Tobacco addiction. *Lancet* 371(9629), 2027–2038 (2008).
- 6 LeSage MG, Keyler DE, Pentel PR. Current status of immunologic approaches to treating tobacco dependence: vaccines and nicotine-specific antibodies. *AAPS J.* 8(1), E65–E75 (2006).
- 7 Gentry WB, Rüedi-Bettschen D, Owens SM. Development of active and passive human vaccines to treat methamphetamine addiction. *Hum. Vaccin.* 5(4), 206–213 (2009).
- 8 Martell BA, Mitchell E, Poling J, Gonsai K, Kosten TR. Vaccine pharmacotherapy for the treatment of cocaine dependence. *Biol. Psychiatry* 58(2), 158–164 (2005).
- 9 Hamaoka T, Takatsu K, Kitagawa M. Antibody production in mice. V. The suppressive effect of anti-carrier antibodies on cellular cooperation in the induction of secondary anti-hapten antibody responses. *Immunology* 24(3), 409–424 (1973).
- 10 Hicks MJ, De BP, Rosenberg JB *et al.* Cocaine analog coupled to disrupted adenovirus: a vaccine strategy to evoke high-titer immunity against addictive drugs. *Mol. Ther.* 19(3), 612–619 (2011).
- 11 Fox BS, Kantak KM, Edwards MA *et al.* Efficacy of a therapeutic cocaine vaccine in rodent models. *Nat. Med.* 2(10), 1129–1132 (1996).
- 12 Bonese KF, Wainer BH, Fitch FW, Rothberg RM, Schuster CR. Changes in heroin self-administration by a rhesus monkey after morphine immunisation. *Nature* 252(5485), 708–710 (1974).
- 13 Peterson EC, Gunnell M, Che Y *et al.* Using hapten design to discover therapeutic monoclonal antibodies for treating methamphetamine abuse. *J. Pharmacol. Exp. Ther.* 322(1), 30–39 (2007).
- 14 McMillan DE, Hardwick WC, Li M *et al.* Effects of murine-derived anti-methamphetamine monoclonal antibodies on (+)-methamphetamine self-administration in the rat. *J. Pharmacol. Exp. Ther.* 309(3), 1248–1255 (2004).
- 15 Roiko SA, Harris AC, LeSage MG, Keyler DE, Pentel PR. Passive immunization with a nicotine-specific monoclonal antibody decreases brain nicotine levels but does not precipitate withdrawal in nicotine-dependent rats. *Pharmacol. Biochem. Behav.* 93(2), 105–111 (2009).
- 16 Keyler D, Pentel PR, Kuehl G, Collins G, Murphy SE. Effects of nicotine infusion on the metabolism of the tobacco carcinogen 4-(methylnitrosamino)-1-(3-pyridyl)-1-butanone (NNK) in rats. *Cancer Lett.* 202(1), 1–9 (2003).
- 17 Norman AB, Tabet MR, Norman MK, Buesing WR, Pesce AJ, Ball WJ. A chimeric human/murine anticocaine monoclonal antibody inhibits the distribution of cocaine to the brain in mice. *J. Pharmacol. Exp. Ther.* 320(1), 145–153 (2007).
- 18 Hieda Y, Keyler DE, Vandervoort JT *et al.* Active immunization alters the plasma nicotine concentration in rats. *J. Pharmacol. Exp. Ther.* 283(3), 1076–1081 (1997).
- 19 Byrnes-Blake KA, Laurenzana EM, Carroll FI *et al.* Pharmacodynamic mechanisms of monoclonal antibody-based antagonism of (+)-methamphetamine in rats. *Eur. J. Pharmacol.* 461(2–3), 119–128 (2003).
- 20 Byrnes-Blake KA, Laurenzana EM, Landes RD, Gentry WB, Owens SM. Monoclonal IgG affinity and treatment time alters antagonism of (+)-methamphetamine effects in rats. *Eur. J. Pharmacol.* 521(1–3), 86–94 (2005).
- 21 Samaha AN, Li Y, Robinson TE. The rate of intravenous cocaine administration determines susceptibility to sensitization. *J. Neurosci.* 22(8), 3244–3250 (2002).
- 22 Gossop M, Griffiths P, Powis B, Strang J. Severity of dependence and route of administration of heroin, cocaine and amphetamines. *Br. J. Addict.* 87(11), 1527–1536 (1992).
- 23 Kosten TR, Rosen M, Bond J *et al.* Human therapeutic cocaine vaccine: safety and immunogenicity. *Vaccine* 20(7–8), 1196–1204 (2002).
- 24 Cornuz J, Zwahlen S, Jungi WF *et al.* A vaccine against nicotine for smoking cessation: a randomized controlled trial. *PLoS ONE* 3(6), e2547 (2008).
- 25 Hatsukami DK, Rennard S, Jorenby D *et al.* Safety and immunogenicity of a nicotine conjugate vaccine in current smokers. *Clin. Pharmacol. Ther.* 78(5), 456–467 (2005).
- 26 Orson F, Rossen RD, Shen X, Lopez AY, Wu Y, Kosten T. Spontaneous development of IgM anti-cocaine antibodies in habitual cocaine users: effect on IgG antibody responses to a cocaine cholera toxin B conjugate vaccine. *Am. J. Addict.* 22(2), 169–174 (2012).
- 27 Norman AB, Norman MK, Buesing WR, Tabet MR, Tsibulsky VL, Ball WJ. The effect of a chimeric human/murine anti-cocaine monoclonal antibody on cocaine self-administration in rats. *J. Pharmacol. Exp. Ther.* 328(3), 873–881 (2009).
- **Early demonstration of the potential and limitations of antibody-based interception of addictive stimulants.**
- 28 Laurenzana EM, Byrnes-Blake KA, Milesi-Hallé A, Gentry WB, Williams DK, Owens SM. Use of anti-(+)-methamphetamine monoclonal antibody to significantly alter (+)-methamphetamine and (+)-amphetamine disposition in rats. *Drug Metab. Dispos.* 31(11), 1320–1326 (2003).
- 29 Raper SE, Chirmule N, Lee FS *et al.* Fatal systemic inflammatory response syndrome in a ornithine transcarbamylase deficient patient following adenoviral gene transfer. *Mol. Genet. Metab.* 80(1–2), 148–158 (2003).
- 30 Lozier JN, Csako G, Mondoro TH *et al.* Toxicity of a first-generation adenoviral vector in rhesus macaques. *Hum. Gene Ther.* 13(1), 113–124 (2002).
- 31 Rivat C, Santilli G, Gaspar HB, Thrasher AJ. Gene therapy for primary immunodeficiencies. *Hum. Gene Ther.* 23(7), 668–675 (2012).
- 32 Stieger K, Lorenz B. Gene therapy for vision loss – recent developments. *Discov. Med.* 10(54), 425–433 (2010).
- 33 Shashidharamurthy R, Bozeman EN, Patel J, Kaur R, Meganathan J, Selvaraj P. Immunotherapeutic strategies for cancer treatment: a novel protein transfer approach for cancer vaccine development. *Med. Res. Rev.* 32(6), 1197–1219 (2012).
- 34 Fang J, Qian JJ, Yi S *et al.* Stable antibody expression at therapeutic levels using the 2A peptide. *Nat. Biotechnol.* 23(5), 584–590 (2005).

- 35 Sanhadji K, Grave L, Touraine JL *et al.* Gene transfer of anti-gp41 antibody and CD4 immunoadhesin strongly reduces the HIV-1 load in humanized severe combined immunodeficient mice. *AIDS* 14(18), 2813–2822 (2000).
- 36 Hicks MJ, Rosenberg JB, De BP *et al.* AAV-directed persistent expression of a gene encoding anti-nicotine antibody for smoking cessation. *Sci. Transl. Med.* 4(140), 140ra87 (2012).
- **Impressive evidence for sustained high-level expression of an extremely effective monoclonal antibody by gene transfer.**
- 37 Russell MA, Wilson C, Patel UA, Feyerabend C, Cole PV. Plasma nicotine levels after smoking cigarettes with high, medium, and low nicotine yields. *Br. Med. J.* 2(5968), 414–416 (1975).
- 38 Rosenberg JB, Hicks MJ, De BP *et al.* AAVrh.10-mediated expression of an anti-cocaine antibody mediates persistent passive immunization that suppresses cocaine-induced behavior. *Hum. Gene Ther.* 23(5), 451–459 (2012).
- 39 Zufferey R, Dull T, Mandel RJ *et al.* Self-inactivating lentivirus vector for safe and efficient *in vivo* gene delivery. *J. Virol.* 72(12), 9873–9880 (1998).
- 40 Haccin-Bey-Abina S, Von Kalle C, Schmidt M *et al.* LMO2-associated clonal T cell proliferation in two patients after gene therapy for SCID-X1. *Science* 302(5644), 415–419 (2003).
- 41 Crystal RG. Transfer of genes to humans: early lessons and obstacles to success. *Science* 270(5235), 404–410 (1995).
- 42 Parks RJ, Chen L, Anton M, Sankar U, Rudnicki MA, Graham FL. A helper-dependent adenovirus vector system: removal of helper virus by Cre-mediated excision of the viral packaging signal. *Proc. Natl Acad. Sci. USA* 93(24), 13565–13570 (1996).
- 43 Kaplitt MG, Leone P, Samulski RJ *et al.* Long-term gene expression and phenotypic correction using adeno-associated virus vectors in the mammalian brain. *Nat. Genet.* 8(2), 148–154 (1994).
- 44 Kuntz E, Yusenko MV, Nagy A, Kovacs G. Oligoarray comparative genomic hybridization of renal cell tumors that developed in patients with acquired cystic renal disease. *Hum. Pathol.* 41(9), 1345–1349 (2010).
- 45 Spalding KL, Bhardwaj RD, Buchholz BA, Druid H, Frisén J. Retrospective birth dating of cells in humans. *Cell* 122(1), 133–143 (2005).
- 46 Muruve DA. The innate immune response to adenovirus vectors. *Hum. Gene Ther.* 15(12), 1157–1166 (2004).
- 47 Brunetti-Pierri N, Ng P. Progress and prospects: gene therapy for genetic diseases with helper-dependent adenoviral vectors. *Gene Ther.* 15(8), 553–560 (2008).
- **A sobering overview of the risks and obstacles associated with adenoviral gene transfer in primates and humans.**
- 48 Brunetti-Pierri N, Ng P. Helper-dependent adenoviral vectors for liver-directed gene therapy. *Hum. Mol. Genet.* 20(R1), R7–13 (2011).
- 49 Morral N, O'Neal W, Rice K *et al.* Administration of helper-dependent adenoviral vectors and sequential delivery of different vector serotype for long-term liver-directed gene transfer in baboons. *Proc. Natl Acad. Sci. USA* 96(22), 12816–12821 (1999).
- 50 Brantly ML, Chulay JD, Wang L *et al.* Sustained transgene expression despite T lymphocyte responses in a clinical trial of rAAV1-AAT gene therapy. *Proc. Natl Acad. Sci. USA* 106(38), 16363–16368 (2009).
- 51 Manno CS, Pierce GF, Arruda VR *et al.* Successful transduction of liver in hemophilia by AAV-Factor IX and limitations imposed by the host immune response. *Nat. Med.* 12(3), 342–347 (2006).
- 52 Vetrini F, Ng P. Liver-directed gene therapy with helper-dependent adenoviral vectors: current state of the art and future challenges. *Curr. Pharm. Des.* 17(24), 2488–2499 (2011).
- 53 Seregin SS, Appledorn DM, McBride AJ *et al.* Transient pretreatment with glucocorticoid ablates innate toxicity of systemically delivered adenoviral vectors without reducing efficacy. *Mol. Ther.* 17(4), 685–696 (2009).
- **A readily applicable means for increasing the success of viral gene transfer while reducing toxicity.**
- 54 Kumahara K, Nagata H, Watanabe K *et al.* Suppression of inflammation by dexamethasone prolongs adenoviral vector-mediated transgene expression in murine nasal mucosa. *Acta Otolaryngol.* 125(12), 1301–1306 (2005).
- 55 Brunetti-Pierri N, Palmer DJ, Beaudet AL, Carey KD, Finegold M, Ng P. Acute toxicity after high-dose systemic injection of helper-dependent adenoviral vectors into nonhuman primates. *Hum. Gene Ther.* 15(1), 35–46 (2004).
- 56 Stewart DJ, Inaba T, Tang BK, Kalow W. Hydrolysis of cocaine in human plasma by cholinesterase. *Life Sci.* 20(9), 1557–1563 (1977).
- 57 Howard LA, Sellers EM, Tyndale RF. The role of pharmacogenetically-variable cytochrome P450 enzymes in drug abuse and dependence. *Pharmacogenomics* 3(2), 185–199 (2002).
- 58 Sellers EM, Kaplan HL, Tyndale RF. Inhibition of cytochrome P450 2A6 increases nicotine's oral bioavailability and decreases smoking. *Clin. Pharmacol. Ther.* 68(1), 35–43 (2000).
- 59 Gorelick DA. Enhancing cocaine metabolism with butyrylcholinesterase as a treatment strategy. *Drug Alcohol Depend.* 48(3), 159–165 (1997).
- 60 Xie W, Altamirano CV, Bartels CF, Speirs RJ, Cashman JR, Lockridge O. An improved cocaine hydrolase: the A328Y mutant of human butyrylcholinesterase is 4-fold more efficient. *Mol. Pharmacol.* 55(1), 83–91 (1999).
- 61 Sun H, Pang YP, Lockridge O, Brimijoin S. Re-engineering butyrylcholinesterase as a cocaine hydrolase. *Mol. Pharmacol.* 62(2), 220–224 (2002).
- 62 Sun L, Lau CE. Simultaneous pharmacokinetic modeling of cocaine and its metabolites, norcocaine and benzyoilecgonine, after intravenous and oral administration in rats. *Drug Metab. Disp.* 29, 1183–1189 (2001).
- 63 Pan Y, Gao D, Yang W *et al.* Computational redesign of human butyrylcholinesterase for anticocaine medication. *Proc. Natl Acad. Sci. USA* 102(46), 16656–16661 (2005).
- 64 Yang W, Xue L, Fang L, Chen X, Zhan CG. Characterization of a high-activity mutant of human butyrylcholinesterase against (-)-cocaine. *Chem. Biol. Interact.* 187(1–3), 148–152 (2010).
- 65 Zheng F, Yang W, Ko MC *et al.* Most efficient cocaine hydrolase designed by virtual screening of transition states. *J. Am. Chem. Soc.* 130(36), 12148–12155 (2008).
- 66 Brimijoin S, Gao Y, Anker JJ *et al.* A cocaine hydrolase engineered from human butyrylcholinesterase selectively blocks cocaine toxicity and reinstatement of drug seeking in rats. *Neuropsychopharmacology* 33(11), 2715–2725 (2008).
- 67 Anker JJ, Brimijoin S, Gao Y *et al.* Cocaine hydrolase encoded in viral vector blocks the reinstatement of cocaine seeking in rats for 6 months. *Biol. Psychiatry* 71(8), 700–705 (2012).

- Shows that gene transfer of engineered cocaine hydrolase can provide quasi-permanent suppression of cocaine-primed drug seeking.
- 68 Saxena A, Sun W, Fedorko JM, Koplovitz I, Doctor BP. Prophylaxis with human serum butyrylcholinesterase protects guinea pigs exposed to multiple lethal doses of soman or VX. *Biochem. Pharmacol.* 81(1), 164–169 (2011).
- 69 Saxena A, Sun W, Luo C, Doctor BP. Human serum butyrylcholinesterase: *in vitro* and *in vivo* stability, pharmacokinetics, and safety in mice. *Chem. Biol. Interact.* 199–203 (2005).
- 70 Weber A, Butterweck H, Mais-Paul U *et al.* Biochemical, molecular and preclinical characterization of a double-virus-reduced human butyrylcholinesterase preparation designed for clinical use. *Vox Sang.* 100(3), 285–297 (2011).
- 71 Gao Y, Atanasova E, Sui N, Pancook JD, Watkins JD, Brimijoin S. Gene transfer of cocaine hydrolase suppresses cardiovascular responses to cocaine in rats. *Mol. Pharmacol.* 67(1), 204–211 (2005).
- 72 Mitani K, Graham FL, Caskey CT, Kochanek S. Rescue, propagation, and partial purification of a helper virus-dependent adenovirus vector. *Proc. Natl Acad. Sci. USA* 92(9), 3854–3858 (1995).
- 73 Kochanek S, Clemens PR, Mitani K, Chen HH, Chan S, Caskey CT. A new adenoviral vector: Replacement of all viral coding sequences with 28 kb of DNA independently expressing both full-length dystrophin and β -galactosidase. *Proc. Natl Acad. Sci. USA* 93(12), 5731–5736 (1996).
- 74 Miyagi N, Rao VP, Ricci D *et al.* Efficient and durable gene transfer to transplanted heart using adeno-associated virus 9 vector. *J. Heart Lung Transplant.* 27(5), 554–560 (2008).
- 75 Schirmer JM, Miyagi N, Rao VP *et al.* Recombinant adeno-associated virus vector for gene transfer to the transplanted rat heart. *Transpl. Int.* 20(6), 550–557 (2007).
- 76 Kim IH, Józkwicz A, Piedra PA, Oka K, Chan L. Lifetime correction of genetic deficiency in mice with a single injection of helper-dependent adenoviral vector. *Proc. Natl Acad. Sci. USA* 98(23), 13282–13287 (2001).
- 77 Gao Y, Geng L, Orson F *et al.* Effects of anti-cocaine vaccine and viral gene transfer of cocaine hydrolase in mice on cocaine toxicity including motor strength and liver damage. *Chem Biol. Interact.* doi:10.1016/j.cbi.2012.08.006 (2012) (Epub ahead of print).
- 78 Wise RA, Kiyatkin EA. Differentiating the rapid actions of cocaine. *Nat. Rev. Neurosci.* 12(8), 479–484 (2011).
- Trenchant analysis of the importance of interoceptive cues in triggering and maintaining drug-seeking behavior.
- 79 Ramakrishnan M, Alves De Melo F, Kinsey BM, Ladbury JE, Kosten TR, Orson FM. Probing cocaine-antibody interactions in buffer and human serum. *PLoS ONE* 7(7), e40518 (2012).
- 80 Gao Y, Orson FM, Kinsey B, Kosten T, Brimijoin S. The concept of pharmacologic cocaine interception as a treatment for drug abuse. *Chem. Biol. Interact.* 187(1–3), 421–424 (2010).
- 81 Carroll ME, Zlebnik NE, Anker JJ *et al.* Combined cocaine hydrolase gene transfer and anti-cocaine vaccine synergistically block cocaine-induced locomotion. *PLoS ONE* 7(8), e43536 (2012).
- 82 Mastrangeli A, Harvey BG, Yao J *et al.* ‘Sero-switch’ adenovirus-mediated *in vivo* gene transfer: circumvention of anti-adenovirus humoral immune defenses against repeat adenovirus vector administration by changing the adenovirus serotype. *Hum. Gene Ther.* 7(1), 79–87 (1996).
- 83 Gao Y, Brimijoin S. Lasting reduction of cocaine action in neostriatum—a hydrolase gene therapy approach. *J. Pharmacol. Exp. Ther.* 330(2), 449–457 (2009).
- 84 Scott EM. Inheritance of two types of deficiency of human serum cholinesterase. *Ann. Hum. Genet.* 37(2), 139–143 (1973).
- 85 Hodgkin W, Giblett ER, Levine H, Bauer W, Motulsky AG. Complete pseudocholesterase deficiency: genetic and immunologic characterization. *J. Clin. Invest.* 44, 486–493 (1965).
- 86 Xue L, Ko MC, Tong M *et al.* Design, preparation, and characterization of high-activity mutants of human butyrylcholinesterase specific for detoxification of cocaine. *Mol. Pharmacol.* 79(2), 290–297 (2011).
- 87 Myers TM, Sun W, Naik RS, Clark MG, Doctor BP, Saxena A. Characterization of human serum butyrylcholinesterase in rhesus monkeys: behavioral and physiological effects. *Neurotoxicol. Teratol.* 34(3), 323–330 (2012).
- 88 Rosenberg YJ, Saxena A, Sun W *et al.* Demonstration of *in vivo* stability and lack of immunogenicity of a polyethyleneglycol-conjugated recombinant CHO-derived butyrylcholinesterase bioscavenger using a homologous macaque model. *Chem. Biol. Interact.* 187(1–3), 279–286 (2010).
- 89 Genovese RF, Sun W, Johnson CC, Ditargiani RC, Doctor BP, Saxena A. Safety of administration of human butyrylcholinesterase and its conjugates with soman or VX in rats. *Basic Clin. Pharmacol. Toxicol.* 106(5), 428–434 (2010).
- 90 Li B, Stribley JA, Ticu A *et al.* Abundant tissue butyrylcholinesterase and its possible function in the acetylcholinesterase knockout mouse. *J. Neurochem.* 75(3), 1320–1331 (2000).
- 91 Silver A. *Biology of Cholinesterases*. North Holland Publishing Co., Amsterdam, The Netherlands (1974).
- 92 Anglister L, Eichler J, Szabo M, Haesaert B, Salpeter MM. 125I-labeled fasciculin 2: a new tool for quantitation of acetylcholinesterase densities at synaptic sites by EM-autoradiography. *J. Neurosci. Methods* 81(1–2), 63–71 (1998).
- 93 Duysen EG, Lockridge O. Whole body and tissue imaging of the butyrylcholinesterase knockout mouse injected with near infrared dye labeled butyrylcholinesterase. *Chem. Biol. Interact.* 175(1–3), 119–124 (2008).

Website

- 101 Painter D, Tice P. Results from the 2010 National Survey on Drug Use and Health: Summary of National Findings (2010). <http://oas.samhsa.gov/NSDUH/2k10NSDUH/2k10Results.pdf>